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## SYNTHESIS AND ANTIVIRAL ACTIVITY OF DIHYDROXYCYCLOHEXYL PYRIMIDINE AND PURINE CARBOCYCLIC NUCLEOSIDES

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**Abstract**: Some novel dihydroxycyclohexyl pyrimidine and purine carbocyclic nucleosides were prepared and tested for inhibitory activity against Cytomegalovirus and Herpes virus-1 and -2.

The pathologies related to viral and/or neoplastic affections are one of the primary causes of death in the world. Natural nucleosides have been the primary models for the design of the most important antitumoral and/or antiviral antimetabolites. The difference which these last two classes of compounds exhibit compared with reference molecules concerns both the basic heterocyclic portion and the glucidic one. The broad majority of therapeutic nucleosides contain a glycosidic moiety. It has been found that its substitution with a carbocyclic

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structure including several hydroxy groups leads to compounds sometimes possessing interesting pharmacological profiles.<sup>3-8</sup> The carbocyclic system most frequently used is the cyclopentane one, chosen because of its structural analogy with the tetrahydrofuran ring present in natural nucleosides.<sup>9,10</sup> More recently, the interesting pharmacological properties of a purine derivative of an oxetane (oxetanocine A, 1) and of its cyclobutane carbanalogues (cyclobut-A, 2 or cyclobut-B, 3) have been reported,<sup>11</sup> these compounds were found to be active towards herpes viruses and HIV, thus indicating that in carbocyclic nucleoside (carbonucleoside) analogues, the cyclic structure was not necessarily a cyclopentane.

On the basis of these data, we decided to prepare some new carbonucleosides in which the carbocyclic structures, different from those already utilized, are linked to pyrimidine (Uracil) or purine (Adenine and Guanine) bases present in pharmacologically active nucleosides.

We therefore synthesized the new cyclohexanyl carbonucleosides<sup>12</sup> of type A (4 and 5) and B (6-11), which present, on the cyclohexane ring, a hydroxy group adjacent and trans with respect to the basic nucleus; this relationship is the same present in natural ribonucleosides and also in several synthetic ones. A and B differ in the position and stereochemistry of the other substituent on the cyclohexane ring.

As starting material for the synthesis of 4-11, we used the cis 12 and trans 13 benzyloxyepoxides.<sup>13</sup> The reaction of the lithium salt of uracil with the cis epoxide 12 at 140 °C in the presence of LiClO<sub>4</sub> afforded only the trans regioisomer 4<sup>14, 15</sup> which was submitted to hydrogenolysis with H<sub>2</sub> in the presence of 10% Pd/C to yield the corresponding diol derivative 5<sup>15</sup> (Scheme 1).

## **SCHEME 1**

Reagents and conditions: (a) Uracil, n-BuLi 1.6M hexane, LiClO 4, DMF, 140 °C 24 h, 30%; (b) H<sub>2</sub>, Pd/C 10%, CH<sub>3</sub>COOH cat., MeOH, 24 h, 65%.

The above-described synthetic conditions were also used for the preparation of the uracil carbocycles  $6^{15}$  and  $7^{15}$ , starting in this case from the trans epoxide 13. In the case of the synthesis of the purine derivatives 8-11, the opening of 13 was carried out at 110 °C using the sodium salt of adenine for  $8^{14.15}$  or the sodium salt of guanine for  $10^{14.15}$ , generated from the appropriate purinic base with NaH in DMF. Hydrogenolysis of benzyl derivatives 8 and 10 with  $H_2$  in the presence of 10% Pd/C gave the desired carbonucleosides  $9^{15}$  and  $11^{15}$  (Scheme 2).

The antiviral activity of the compounds synthesized was evaluated by the plaque-reduction method against Cytomegalovirus (CMV) in PEU cells and Herpes virus 1 (HSV-1) and 2 (HSV-2), in Vero cells<sup>17,18</sup>. Results are shown in Table 1. The carbonucleosides 4, 5 and 7 exhibited quite a good efficacy against the three virus species tested, while the carbonucleosides 8, 9 and 10 were effective only against HSV-1 and -2. No cytotoxicity was observed for any of the compounds.

## **SCHEME 2**

Reagents and conditions: (a) Uracil, n-BuLi 1.6 M hexane, LiClO 4, DMF, 140 °C, 24h, 34% H<sub>2</sub>, Pd/C 10%, CH<sub>3</sub>COOH cat., MeOH, 24 h, 65%; (c) Adenine or Guanine, NaH, DMF, 110 °C, 24 h, 20% for Aden. or 23% for Guan.; (d) H<sub>2</sub>, Pd/C 10%, CH<sub>3</sub>COOH, 48 h.

Table 1. Antiviral activity and cytotoxicity of the carbonucleosides.

Compound	EC <sub>50</sub> (μM) <sup>a</sup>			IC 50 (μM) <sup>b</sup>
	CMV	HSV-1	HSV-2	PEU cells
4	10.0	10.0	10.0	>250
5	0 97	10 0	0.97	>250
6	10 0	158	158	>250
7	10.0	0 97	0.97	>250
8	147	16.8	16.2	>250
9	200	28.5	30.0	>250
10	140	21 2	22.1	>250
11	188	188	188	>250

<sup>&</sup>lt;sup>a</sup>Concentration required to inhibit CMV-, HSV-1- or HSV-2-induced cytopathic effects by 50 %. <sup>b</sup>The lowest concentration of the compound producing overt cytotoxicity in virus-free cultures.

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- 12. Recently new nucleosides different from ours in which the glucidic portion has been substituted with a cyclohexanol nucleus have been reported in literature. See for example: [a] Ramesh,K.; Wolfe, M.S.; Lee, Y.; Velde, D.V.; Brochardt, R.T. J. Org. Chem. 1992, 57, 5861. [b] Jähne, G.; Winkler, I.; Helsberg, M., Hänel, H. European Patent Application No. 92111063.1 filed June 30, 1992. [c] Shiozawa, A.; Matsubara K.; Nagata, T.; Hoshino, H.; Saki, J. European Patent Application No. 91111925.3 filed July 17, 1991. [d] Arango, J.H.; Geer, A.; Rodriguez, J.; Young, P.E.; Scheiner, P. Nucleosides and Nucleotides 1993, 12, 773.
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- 14. The trans relationship between the OH and the basic heterocyclic portion in the addition products (4, 6, 8 and 10) of epoxides 12 and 13 with pyrimidinic or purinic bases was assumed on the basis of the usual anti additions to these epoxide systems. The regiochemistry of the addition products (4, 6, 8 and 10) was assumed on the basis of findings for ring opening reactions of 12 and 13 with nucleophiles. However the exact structure and regiochemistry of the benzyl derivative 4 was unequivocally determined by X-ray analysis (material available from our laboratory). The site of alkylation (N-9) for the purine derivatives (8-11) was determined by UV spectroscopy: data obtained for 9 and 11 were in close agreement with those found for the natural purinic nucleosides.

- 15. All compounds were well characterized spectroscopically. 4 m.p. 237 °C, ¹HNMR (200 MHz, DMSOd<sub>6</sub>)  $\delta$  11.13 (s, 1H), 7.69 (d, J=8.0 Hz, 1H), 7.22-7.36 (m, 5H), 5.54 (d, J=8.0 Hz, 1H), 4.92 (m, 1H), 4.39-4.51 (m, 3H), 3.62-3.74 (br, 2H), 1.37-2.01 (m, 6H). 5: m.p. 247 °C; <sup>1</sup>HNMR (200 MHz, DMSOd<sub>6</sub>) δ 11.09 (s, 1H), 7.66 (d, J=8.0 Hz, 1H), 5.51 (d, J=8.0 Hz, 1H), 4.84 (m, 1H), 4.42-4.62 (m, 2H), 3.88-3.97 (m, 1H), 3.61 (br, 1H), 1.44-1.67 (m, 6H), 6; m.p. 189 °C; ¹HNMR (200 MHz, DMSO-d<sub>6</sub>) δ 11.15 (s, 1H), 7.54 (d, J=8.0 Hz, 1H), 7.23-7.38 (m, 5H), 5.52 (d, J=8.0 Hz, 1H), 4.91 (m, 1H), 4.40-4.53 (m, 2H), 3.91-4.09 (m, 2H), 3.75 (s, 1H), 1.37-2.24 (m, 6H). 7: m.p. 189 °C; <sup>1</sup>HNMR (200 MHz, DMSO-d<sub>6</sub>)  $\delta$  11.13 (s, 1H), 7.56 (d, J=8.0 Hz, 1H), 5.53 (d, J=8.0 Hz, 1H), 4.81 (m, 1H), 4.55 (m, 1H) 3.86-4.07 (m, 3H), 1.33-2.03 (m, 6H). 8: m.p. 262 °C; <sup>1</sup>HNMR (200 MHz, DMSO-d<sub>6</sub>) δ 8.10 (s, 2H), 7.24-7.45 (m, 5H), 7.12 (s, 2H), 4.83 (m, 1H), 4.45-4.58 (m, 2H), 4.28-4.40 (m, 1H), 4.07-4.20 (m, 1H), 3.82 (br, 1H), 1.43-2.45 (m, 6H). 9: m.p. 258 °C,  $^{1}$ HNMR (200 MHz, DMSO-d<sub>6</sub>)  $\delta$  8.08 (s, 2H), 7.10 (s, 2H), 4.73 (m, 1H), 4.61 (br, 1H), 4.28-4.36 (m, 1H), 4.01-4.14 (m, 2H), 1.39-2.43 (m, 6H), UV  $\lambda_{\text{max}}$  (MeOH) sh 261 nm ( $\epsilon$  13300). 10: m.p. 295 °C dec.; <sup>1</sup>HNMR (200 MHz, DMSO-d<sub>6</sub>)  $\delta$  10.52 (brs, 1H), 7.71 (s, 1H), 7.25-7.38 (m, 5H), 6.34 (s, 2H), 4.87 (m, 1H), 4.44-4.57 (m, 2H), 3.97-4.17 (m, 2H), 3.79 (br, 1H), 1.39-2.28 (m, 6H). 11: m.p. 290°C dec.; <sup>1</sup>HNMR (200 MHz, DMSO- $d_0$ )  $\delta$  10.56 (brs, 1H), 7.67 (s, 1H), 6.54 (s, 2H), 4.76 (br, 1H), 4.12-4.16 (m, 1H), 3.85-3.99 (m, 3H), 1.34-2.12 (m, 6H); UV  $\lambda_{max}$  (MeOH) sh 254 nm ( $\epsilon$  12280).
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